

Determination of the Fatty Acid Composition of the *Dasyatis sephen* (F.) Liver Oil Captured from Cost of Jaffna Sri Lanka

DUGLAS SATHEES

<https://orcid.org/0000-0001-8873-5148>

duglassathees@gmail.com

Department of Animal Science,
Faculty of Agriculture, University of Jaffna
Sri Lanka

VIDANARACHCHI J.K

Department of Animal Science
Faculty of Agriculture, University of Peradeniya
Sri Lanka

HIMALI S.M.C

Department of Animal Science
Faculty of Agriculture, University of Peradeniya
Sri Lanka

ABSTRACT

The liver oil from the cartilaginous fish was highly composed of poly- and highly unsaturated fatty acids. Extraction of oil from the *Dasyatis sephen* liver is simple and cheap. Therefore, the present study was conducted to investigate proximate Lipid Profile composition and physicochemical properties of *Dasyatis sephen* liver oil. Gas-Liquid Chromatography method

(GLC) was used to determine the Lipid Fatty acids profiles. The average Lipid level of *Dasyatis sephen* liver oil was found to be 69.54 % (w/w), greater than that from the *D. pastinaca* (58.27%) and *D. violacea* (57.33%). Crude liver lipid content was of were highly significant. The total SFA percentages was 44.2% and the predominant was C16 (palmitic acid) about 35.0%. Surprisingly, unsaturated fatty acids profiles of 20:5n-3 and 22:6n-3 exhibited as 0.5 % and 0.6 %. Physio-chemical properties such as moisture content, color, specific gravity, peroxide value, and fatty acid compositions were obtained under the tolerable standard. It demonstrated one of the locally available resources currently being wasted has the potential to use in the manufacturing of pharmaceutical and nutraceutical industries.

KEYWORDS Fish Liver Oil, *Dasyatis sephen*, Saturated fatty acids, Unsaturated fatty acids, Sri Lanka

INTRODUCTION

Lipid fraction extracted from tissues of oily fish and fishery by-products is known as Fish oil. It is one of the good sources of polyunsaturated fatty (PUFAs) Omega-3 (ω -3) such as eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA). This paper elaborates the study performed on the *Dasyatis sephen* liver oils from the dry fish processing industry were usually directly discarded into the sea.

Ray fish in Jaffna of Sri Lanka represent an interesting source of locally available quality fish. In this region, *Dasyatis sephenes* are caught for meat and dry fish production. Typically removal of the pectoral fins and discarding the remaining of the body eviscerates directly discarded into the sea. Nevertheless, it is well known that the Liver of *Dasyatis sephen* is rich in poly- and highly unsaturated fatty acids (PUFA and HUFA, respectively). It has been estimated that over 15000 kg of Ray fish is caught in Northern Sri Lanka each month (Department of Fisheries Report 2016).

Though Fish oil has numerous health benefits, only few can afford to take it because it is much expensive. Fluctuating market prices for Fish also have been an issue in fish oil production. Recently, the production of fish oil is becoming more demanding, as there is a sizable growing demand for high-quality fish oil in the world market.

Apart from its use as edible oil, it also has functional benefits in both pharmaceuticals and industries. A literature study done in the past has shown that little work has been done in terms of *Dasyatis sephen* liver oil and its fish by-products. Therefore, *Dasyatis sephen* liver oil may have used

as commercial raw materials for the manufacturing of pharmaceutical and nutraceutical products.

METHODOLOGY

Study Location

The study was carried out in the Gurunagar and Delft Islands Fisheries Harbors of Jaffna, Northern Provenience in Sri Lanka. All the Analytical tests were done in the Nutrition Laboratory and Meat Science Laboratory, Department of Animal Science, Food Science Laboratory, Department of Food Science, Faculty of Agriculture, University of Peradeniya, and in the Beuro Venters, Colombo, Sri Lanka.



(Dorsal View)

Plate 2.1: Sting Ray (*Dasyatis sephen*)

Sample Collection and Preparation

Three samples of *Dasyatis sephen* Fish (Forsskal, 1175) were collected from each Fisheries Harbors and Dry Fish Cottages of Jaffna, Northern Provenience in Sri Lanka. Samples were transported in sealed freezer containers to laboratory at 0 °C. They were stored at (-20 °C) in the deep freezer for analytical purposes. Each sample was taken out to thaw at room temperature to measure its lengths and weights as a whole. Samples' livers, offal (gut & tail), and carcass were eviscerated, and stored at -20 °C in sealed poly bags separately for further analysis.

Hepatosomatic Index (HIS)

The liver weight to body weight ration was calculated.

Extraction of *Dasyatis sephen* Liver Crude Oil

Frozen livers were weighted and thawed at room temperature at 27 °C. The livers were washed and cleaned. Then they were sliced to obtain steaks. The liver steaks were minced thoroughly using the electric homogenizer. The Homogenized samples were weighted.

The modified Bligh and Dyer method (1959) was used to extract the crude oil in the liver samples. 50 g liver samples were homogenized for two minutes using an electric homogenizer with 100 mL of methanol and 50 mL of chloroform. Then, again 50-mL of chloroform was added to homogenize for another 30 Seconds. The homogenized mixture was diluted with 50 mL of distilled water.

Mixture samples were filtered by a Whatman No.42 filter paper lined with Buncher funnel under vacuum suction. 20 mL of chloroform used to rinse the residues, and the filtered portion was transferred to a separatory funnel. The chloroform layer containing liver oil (bottom layer) was separated by removing it. Whatman No.1 filter paper lined with Buncher funnel under vacuum suction. It was used to Filter Finely. 20 mL of chloroform used to rinse the residues. To remove moisture, the filtrate was passed through 3 g of anhydrous sodium sulfate.

Extraction of Fish oil from the Chloroform layer was carried out by transferring the filtrate into a dry pre-weighed round-bottom flask of rotary vacuum evaporator. The chloroform was removed using a rotary vacuum evaporator at 40 °C. Extracted crude liver oil was weighed and treated with 0.02 BTH to keep moisture free. The prepared crude liver oil was stored at (-20 °C) for further analysis.

Crude Oil Recovery Percentage

Given below equation was used to calculate the Crude oil content in the Liver sample. Equation: Percentage of lipid in the fish liver

$$\% \text{ Lipid} = \frac{W_2}{W_s} \times 100$$

W_s = Weight of the sample (g)

W_2 = Weight of Lipid extracted after evaporation (g)

Determination of fatty acid composition in Crude oil

Gas-Liquid Chromatography (GLC) method was used to determined Fatty acids profiles, according to Buchgraber et al. (2000).

Preparation of fatty acid methyl ester (FAME)

0.074 g of Sodium methoxide (CH_3NaO) (0.5M) was weighed accurately and dissolved in 2 mL of methanol. 100 mg of methylheptadecanoate acid (1mg/mL) was dissolved in 100 ml of hexane.

100 mg sample was weighed into a 15 mL screw-capped methylation tube, and 1 mg/mL of internal standard (Methylheptadecanoate), 2mL of 0.5M sodium methoxide, and 300 UL of dichloromethane were added. Meanwhile, the mixture was kept in a heat block at 50°C for 30min. It was allowed to cool to room temperature. Drop by drop 5mL of distilled water was added. After that, 100 UL of glacial acetic was added.

The tube with contents was centrifuged at the speed of 1500rpm for 10 minutes at 5°C. The top hexane layer was separated and added in to a 2ml GC Vial. Again 500 UL of hexane was added and centrifuged at the speed of 1500rpm for 10 minutes at 5°C. The top hexane layer was separated and added to the same GC vial. Vials were sealed with Para film and frozen immediately at (-20°C) until GC analysis.

Gas-Liquid chromatography analysis of the sample

The prepared FAME samples were analyzed by injecting 1 μL into GLC (Shimadzu, 14-B, Japan), equipped with a Flame Ionization Detector (FID). A fused silica capillary column (100 m, 0.25 mm id and 0.20 μm film thickness) attached with Chromotopac data processor (Model-CR6A, Shimadzu, Japan). The split ratio was 100:1. Temperatures of the Injector and detector were maintained at 260°C. Helium was used as carrier gas at a flow rate of 20 mL/sec. The initial column oven temperature was maintained at 140°C for 5 min and increased to 220°C at the rate of 4°C/min, then maintained at that temperature for 10 min. Fatty acids were identified by comparison of their retention time with authentic standards (SUPELCO 37 Component FAME Mix, Sigma Aldrich) (Kuksis et al., 1967). The amount of each fatty acid were expressed as a percentage (%) of the sum of all fatty acids in the sample.

RESULTS AND DISCUSSION

Hepatosomatic Index (HSI) of *Dasyatis sephen* liver

The average body weights were found to be 2301.05g, 2594.17g, and 2847.84g average lengths were 137.67cm, 153.49cm and 175.74cm. The average liver weight were 253.69g, 289.52g and 313.98 g respectively. Calculated HSI value was 9.05 %. Moreover, the average lengths of *Dasyatis sephen* were measured to be. The HSI value of *Dasyatis sephen* fish is comparatively greater than the findings of Özyılmaz and Öksüz, 2015 which is 8.25 %. This may be due to the increased liver weight and the maturity level of fish.

Table 4.1: The average length (cm), Total weight (g), Liver weight (g) and Hepatosomatic index (HSI) of the *Dasyatis sephen*.

Fish species	Length (cm)	Total weight (g)	Liver weight (g)	HSI (%)
Stingray- ₁	137.67	2301.05	253.69	9.07
Stingray- ₂	153.49	2594.17	289.52	8.96
Stingray- ₃	175.74	2847.84	311.92	9.13

Mean±SD values were presented (n = 3)

Dasyatis sephen Liver Crude Oil Recovery Percentage

The average liver crude oil recovery of the *asyatis sephen* was 69.54 %. All of the *Dasyatis sephen* had a large amount of lipids in their liver. However, the levels of liver lipid not varied greatly from one species to another (Özyılmaz and Öksüz, 2015).



Plate: 4.1 Fish oil Colour

Fatty acid Profile of *Dasyatis sephen* crude liver oil

The predominant SFA fatty acid in the stingray was C16 (palmitic acid), which had a value of 10.2%. Clearly, the average level of C16 in the *Dasyatis sephen* liver oil was significantly higher than the level of the other *Dasyatis sephen* liver oils. The total levels of PUFA in all the fish liver oils seems similar.

The total SFA percentages of *Dasyatis sephen* fish in this study was 44.2%. The Total SFA percentages of *Dasyatis sephen* liver oils from *Dasyatis sephen* that were previously reported were 34.97%, respectively (Navarro-Garcia et al., 2010). The percentages of total SFA in *Dasyatis sephen* liver oil were not in agreement with the finding from the current study.

Table 3.1. Fatty acid composition of stingray fish liver oil

Carbon Chain	Stingray
C12:0	4.1
C13:0	0.7
C14:00	0.3
C16:0	35.0
C17:0	0.4
C18:0	2.5
C20:0	1.2
ΣSFA	44.2
C16:1	10.2
C18:1	1.3
ΣMUFA	11.5
C18:2	0.3
C20:4	1.4
Σn6	1.7
C20:5n3	0.5
C22:6n3	0.6
Σn3	1.1
ΣPUFA	2.8
n3/n6	0.64
DHA/EPA	0.83

CONCLUSIONS

Dasyatis sephen liver was obtained during dry fish processing Cottages of Jaffna, Northern Province, Sri Lanka. Liver oil extracted through the Bligh and Dyer method gave the total recovery percentage of 69.54 % with high quality.

The total SFA percentages was 44.2%, and with predominant C16 (palmitic acid) about 35.0%. Unsaturated fatty acids profiles of 20:5n-3 and 22:6n-3 exhibited as 0.5 % and 0.6 %. Physio-chemical properties such as moisture content, color, specific gravity, peroxide value, and fatty acid compositions were obtained under the tolerable standard. Oil extracted from *Dasyatis sephen* liver eviscerates from the dry fish industry could be a potential source for pharmaceutical and nutraceutical industries.

LITERATURE CITED

- Ackman RG. Fish in Marine Biogenic Lipids, Fats and Oils. CRC Press, Inc., Boca Raton 1989; 2:145-178. Retrieved on April 28, 2017 from <https://bit.ly/3vwgW4g>
- Ackman RG. Nutritional Composition of Fats in Sea foods. Prog Food Nutr Sci 1989; 13:161–241. Retrieved on April 28, 2017 from <https://europepmc.org/article/med/2699043>
- Ackman, R. G. (2006). Marine lipids and omega-3 fatty acids. Handbook of functional lipids, 311-324. Retrieved on April 28, 2017 from <https://www.cabdirect.org/cabdirect/abstract/20053165005>
- Aidos, I., van der Padt, A., Boom, R. M., & Luten, J. B. (2001). Upgrading of maatjes herring byproducts: production of crude fish oil. Journal of Agricultural and Food Chemistry, 49(8), 3697-3704. Retrieved on April 28, 2017 from <https://pubs.acs.org/doi/abs/10.1021/jf001513s>
- Aidos, I., Masbernart-Martinez, S., Luten, J. B., Boom, R. M., & Van Der Padt, A. (2002). Composition and stability of herring oil recovered from sorted byproducts as compared to oil from mixed byproducts. Journal of agricultural and food chemistry, 50(10), 2818-2824. Retrieved on April 28, 2017 from <https://pubs.acs.org/doi/abs/10.1021/jf011318t>
- Aidos, .I, Van Der P.A., Luten, J.B, Boom, R.M. (2002). Seasonal Changes in Crude and Lipid Composition of Herring Fillets, Byproducts, and Respective Produced Oils. Journal of Agricultural and Food Chemistry 16:

- 4589-4599. Retrieved on April 28, 2017 from <https://pubs.acs.org/doi/abs/10.1021/jf0115995>
- Alders, L. (1955). Liquid-Liquid Extraction. Elsevier Publishing. <http://117.239.25.194:7000/jspui/bitstream/123456789/959/1/PRILIMINERY%20AND%20CONTENTS.pdf>
- Barlow, S. M., & Stansby, M. E. (1982). Nutritional evaluation of long-chain fatty acids in fish oil; proceedings. Retrieved on April 28, 2017 from <https://agris.fao.org/agris-search/search.do?recordID=XF2015030091>
- Bimbo, A. P. (1998). Guidelines for characterizing food-grade fish oils. Inform, 9(5). Retrieved on April 28, 2017 from <https://ci.nii.ac.jp/naid/80010360037/>
- Breivik, H. (Ed.). (2007). Long-chain omega-3 specialty oils (Vol. 21). Oily Press. Retrieved on April 28, 2017 from <https://www.sciencedirect.com/book/9780955251214/long-chain-omega-3-specialty-oils>
- Bligh, E. G., & Dyer, W. J. (1959). A rapid method of total lipid extraction and purification. Canadian journal of biochemistry and physiology, 37(8), 911-917. Retrieved on April 28, 2017 from <https://cdnsiencepub.com/doi/abs/10.1139/059-099>
- Boran, G., H. Karacam and M. Boran. 2006. Changes in the quality of fish oils due to storage temperature and time. Food Chemistry, 98, 693-698. Retrieved on April 28, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S0308814605005704>
- Bruinsma, K.A. and D.L. Taren. 2000. Dieting, essential fatty acid intake, and depression, 58(4), 98-108. Retrieved on May 18, 2017 from <https://academic.oup.com/nutritionreviews/article/58/4/98/1857235?login=true>
- Farooqui, A. A. 2009. Beneficial effects of fish oil on human brain. New York: Springer, pp .151-187. Retrieved on May 18, 2017 from <https://link.springer.com/book/10.1007%2F978-1-4419-0543-7>
- Fei, C. Y., Salimon, J., & Said, M. (2010). Optimisation of urea complexation by Box-Behnken design. Sains Malaysiana, 39(5), 795-803. Retrieved on May 18, 2017 from http://www.ukm.my/jsm/pdf_files/SM-PDF-39-5-2010/16%20Chin.pdf
- Frankel, E. N. (1991). Recent advances in lipid oxidation. Journal of the Science of Food and Agriculture, 54(4), 495-511. Retrieved on April 26, 2017 from <https://onlinelibrary.wiley.com/doi/abs/10.1002/jsfa.2740540402>

- Fritsche, K.L. and P.V. Johnston. 1988. Rapid autoxidation of fish oil in diets without added antioxidants. *Journal of Nutrition*, 118, 425-426. Retrieved on April 26, 2017 from <https://academic.oup.com/jn/article-abstract/118/4/425/4737951>
- Gamez-Meza, N., Noriega-Rodriguez, J. A., Medina-Juárez, L. A., Ortega-Garcia, J., Monroy-Rivera, J., Toro-Vázquez, F. J., ... & Angulo-Guerrero, O. (2003). Concentration of eicosapentaenoic acid and docosahexaenoic acid from fish oil by hydrolysis and urea complexation. *Food Research International*, 36(7), 721-727. Retrieved on April 26, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S0963996903000528>
- García, R., Bermejo, C., Grau, C., Pérez, R., Rodríguez-Peña, J. M., Francois, J., ... & Arroyo, J. (2004). The global transcriptional response to transient cell wall damage in *Saccharomyces cerevisiae* and its regulation by the cell integrity signaling pathway. *Journal of Biological Chemistry*, 279(15), 15183-15195. Retrieved on April 26, 2017 from <https://www.sciencedirect.com/science/article/pii/S0021925819639177>
- Gray, J. I., Gomaa, E. A., & Buckley, D. J. (1996). Oxidative quality and shelf life of meats. *Meat science*, 43, 111-123. Retrieved on January 26, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/0309174096000599>
- Haagsma, N., Van Gent, C. M., Luten, J. B., De Jong, R. W., & Van Doorn, E. (1982). Preparation of an ω_3 fatty acid concentrate from cod liver oil. *Journal of the American Oil Chemists' Society*, 59(3), 117-118. Retrieved on January 26, 2017 from <https://aocs.onlinelibrary.wiley.com/doi/abs/10.1007/BF02662254>
- Hayes, D. G., Bengtsson, Y. C., Van Alstine, J. M., & Setterwall, F. (1998). Urea complexation for the rapid, ecologically responsible fractionation of fatty acids from seed oil. *Journal of the American Oil Chemists' Society*, 75(10), 1403-1409. Retrieved on January 26, 2017 from <https://link.springer.com/article/10.1007%2F11746-998-0190-9>
- Helland, I. B., Smith, L., Blomén, B., Saarem, K., Saugstad, O. D., & Drevon, C. A. (2008). Effect of supplementing pregnant and lactating mothers with n-3 very-long-chain fatty acids on children's IQ and body mass index at 7 years of age. *Pediatrics*, 122(2), e472-e479. Retrieved on April 26, 2017 from <https://pediatrics.aappublications.org/content/122/2/e472.short>
- Holub, D. J., & Holub, B. J. (2004). Omega-3 fatty acids from fish oils and

- cardiovascular disease. *Molecular and cellular biochemistry*, 263(1), 217-225. Retrieved on April 26, 2017 from <https://link.springer.com/article/10.1023/B:MCBI.0000041863.11248.8d>
- Huss, H. H. (1988). Fresh fish–quality and quality changes: a training manual prepared for the FAO/DANIDA Training Programme on Fish Technology and Quality Control (No. 29). Food & Agriculture Org. Retrieved on April 26, 2017 from <https://bit.ly/2SeE3ll>
- Iverson, S. J., Lang, S. L., & Cooper, M. H. (2001). Comparison of the Bligh and Dyer and Folch methods for total lipid determination in a broad range of marine tissue. *Lipids*, 36(11), 1283-1287. Retrieved on April 26, 2017 from <https://link.springer.com/article/10.1007/s11745-001-0843-0>
- Iqbal, S., & Bhanger, M. I. (2007). Stabilization of sunflower oil by garlic extract during accelerated storage. *Food Chemistry*, 100(1), 246-254. Retrieved on April 26, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S0308814605008630>
- Jayathilakan, K., Sharma, G. K., Radhakrishna, K., & Bawa, A. S. (2007). Antioxidant potential of synthetic and natural antioxidants and its effect on warmed-over-flavour in different species of meat. *Food Chemistry*, 105(3), 908-916. Retrieved on April 26, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S0308814607003913>
- Lenihan-Geels, G., Bishop, K. S., & Ferguson, L. R. (2013). Alternative sources of omega-3 fats: can we find a sustainable substitute for fish?. *Nutrients*, 5(4), 1301-1315. Retrieved May 7, 2017 from <https://www.mdpi.com/2072-6643/5/4/1301>
- Liu, S., Zhang, C., Hong, P., & Ji, H. (2006). Concentration of docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA) of tuna oil by urea complexation: optimization of process parameters. *Journal of food engineering*, 73(3), 203-209. Retrieved on May 18, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S0260877405000634>
- Kebir, M. V. O. E., Barnathan, G., Gaydou, E. M., Siau, Y., & Miralles, J. (2007). Fatty acids in liver, muscle and gonad of three tropical rays including non-methylene-interrupted dienoic fatty acids. *Lipids*, 42(6), 525-535. Retrieved on February 14, 2017 from <https://aocs.onlinelibrary.wiley.com/doi/abs/10.1007/s11745-007-3040-x>
- Mayneris-Perxachs, J., I. Bondia-Pons, L. Serra-Majem, A.I. Castello, and M.C. López-Sabater (2010). Long-chain n-3 fatty acids and classical

- cardiovascular disease risk factors among the Catalan population. *Food Chem.*, 119: 54-61. Retrieved on February 14, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S030881460900778X>
- Moffat, C. F., & McGill, A. S. (1993). Variability of the composition of fish oils: significance for the diet. *Proceedings of the Nutrition Society*, 52(3), 441-456. Retrieved on February 14, 2017 from <https://www.cambridge.org/core/journals/proceedings-of-the-nutrition-society/article/variability-of-the-composition-of-fish-oils-significance-for-the-diet/D1F47120BED4E559C474F066A4EB72CD>
- Reuber, M. D. (1979). Carcinogenicity of chloroform. *Environmental health perspectives*, 31, 171-182. Retrieved on May 18, 2017 from <https://ehp.niehs.nih.gov/doi/abs/10.1289/ehp.7931171>
- Rizliya, V., & Mendis, E. (2014). Biological, physical, and chemical properties of fish oil and industrial applications. In *Seafood processing by-products* (pp. 285-313). Springer, New York, NY. Retrieved on May 18, 2017 from https://link.springer.com/chapter/10.1007/978-1-4614-9590-1_14
- Navarro-García, G., Ramírez-Suárez, J. C., Cota-Quiñones, E., Márquez-Farías, F., & Bringas-Alvarado, L. (2010). Storage stability of liver oil from two ray (*Rhinoptera bonasus* and *Aetobatus narinari*) species from the Gulf of Mexico. *Food chemistry*, 119(4), 1578-1583. Retrieved on May 18, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S0308814609011054>
- Navarro-García, G., Pacheco-Aguilar, R., Vallejo-Cordova, B., Ramirez-Suarez, J. C., & Bolanos, A. (2000). Lipid composition of the liver oil of shark species from the Caribbean and Gulf of California waters. *Journal of Food Composition and Analysis*, 13(5), 791-798. Retrieved on May 18, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S0889157500909281>
- Navarro-García, G., Pacheco-Aguilar, R., Bringas-Alvarado, L., & Ortega-García, J. (2004). Characterization of the lipid composition and natural antioxidants in the liver oil of *Dasyatis brevis* and *Gymnura marmorata* rays. *Food chemistry*, 87(1), 89-96. May 18, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S0308814603005727>
- Norziah, M.H., J. Nuraini and K.Y. Lee. 2009. Studies on the extraction and characterization of fish oil from wastes of seafood processing industry. *Journal of the Science of Food and Agriculture*, 2(4), 959-973.

- Retrieved on March 28, 2017 from <https://www.cabdirect.org/cabdirect/abstract/20103303573>
- Ozogul, Y., Şimşek, A., Ballıkcı, E., & Kenar, M. (2012). The effects of extraction methods on the contents of fatty acids, especially EPA and DHA in marine lipids. *International journal of food sciences and nutrition*, 63(3), 326-331. Retrieved on March 28, 2017 from <https://www.tandfonline.com/doi/abs/10.3109/09637486.2011.627844>
- Özyılmaz, A., & Öksüz, A. (2015). Determination of the biochemical properties of liver oil from selected cartilaginous fish living in the northeastern Mediterranean. *Journal of Animal and Plant Sciences*, 25, 160-167. Retrieved on May 18, 2017 from <https://bit.ly/3aPV1gF>
- Pak, C. S. (2005). Stability and quality of fish oil during typical domestic application. Fisheries Training Programme, The United Nations University, Iceland. Retrieved on May 18, 2017 from <https://bit.ly/3nGnf2z>
- Pal, D., Banerjee, D., Patra, T. K., Patra, A., & Ghosh, A. (1998). Liver lipids and fatty acids of the sting ray *Dasyatis bleekeri* (Blyth). *Journal of the American Oil Chemists' Society*, 75(10), 1373-1378. Retrieved on May 18, 2017 from <https://link.springer.com/article/10.1007%2Fs11746-998-0185-6>
- Pazos, M., Gallardo, J. M., Torres, J. L., & Medina, I. (2005). Activity of grape polyphenols as inhibitors of the oxidation of fish lipids and frozen fish muscle. *Food Chemistry*, 92(3), 547-557. Retrieved on May 18, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S0308814604006624>
- Pettinello, G., Bertucco, A., Pallado, P., & Stassi, A. (2000). Production of EPA enriched mixtures by supercritical fluid chromatography: from the laboratory scale to the pilot plant. *The Journal of Supercritical Fluids*, 19(1), 51-60. Retrieved on May 18, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S0896844600000723>
- Ramakrishnan, U., Stein, A. D., Parra-Cabrera, S., Wang, M., Imhoff-Kunsch, B., Juárez-Márquez, S., ... & Martorell, R. (2010). Effects of docosahexaenoic acid supplementation during pregnancy on gestational age and size at birth: randomized, double-blind, placebo-controlled trial in Mexico. *Food and nutrition bulletin*, 31(2_suppl2), S108-S116. Retrieved on May 18, 2017 from <https://journals.sagepub.com/doi/abs/10.1177/156482651003125203>
- Ratnayake, W. M. N., Olsson, B., Matthews, D., & Ackman, R. G. (1988).

- Preparation of omega-3 PUFA concentrates from fish oils via urea complexation. *Lipid/Fett*, 90(10), 381-386. Retrieved on May 18, 2017 from <https://onlinelibrary.wiley.com/doi/abs/10.1002/lipi.19880901002>
- Raza, S. A., Rashid, A., William, J., Najaf, S., & Arshad, M. (2009). Effect of synthetic antioxidant on shelf life of locally manufactured butter known as Makhan in Pakistan. *Biharean Biologist*, 3(2), 161-162. Retrieved on February 14, 2017 from <http://www.biozoojournals.ro/bihbiol/cont/v3n2/bb.031205.Raza.pdf>
- Rubio-Rodríguez, N., Sara, M., Beltrán, S., Jaime, I., Sanz, M. T., & Rovira, J. (2008). Supercritical fluid extraction of the omega-3 rich oil contained in hake (*Merluccius capensis*–*Merluccius paradoxus*) by-products: study of the influence of process parameters on the extraction yield and oil quality. *The Journal of Supercritical Fluids*, 47(2), 215-226. Retrieved on February 14, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S089684460800199X>
- Saify, Z. S., Akhtar, S., Khan, K. M., Perveen, S., Ayattollahi, S. A. M., Hassan, S., ... & KHAN, M. Z. (2003). A study on the fatty acid composition of fish liver oil from two marine fish, *Eusphyra blochii* and *Carcharhinus bleekeri*. *Turkish Journal of Chemistry*, 27(2), 251-258. Retrieved on February 14, 2017 from <https://journals.tubitak.gov.tr/chem/abstract.htm?id=6204>
- Sang, W., & Jin, Z. T. (2004). Lipid oxidation of fish liver oil as affected by light, antioxidants and temperature. *Journal of food processing and preservation*, 28(1), 1-10. Retrieved on February 14, 2017 from <https://ifst.onlinelibrary.wiley.com/doi/abs/10.1111/j.1745-4549.2004.tb00533.x>
- Senanayake, S. N., & SHAHIDI, F. (2000). Concentration of docosahexaenoic acid (DHA) from algal oil via urea complexation. *Journal of Food Lipids*, 7(1), 51-61. Retrieved on May 18, 2017 from <https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1745-4522.2000.tb00160.x>
- Simopoulos, A. P. (1991). Omega-3-fatty-acids in health and disease and in growth and development. *American Journal of Clinical Nutrition*, 54(3), 438-463. Retrieved on May 18, 2017 from <https://academic.oup.com/ajcn/article-abstract/54/3/438/4694393>
- Simopoulos, A.P (2006). Evolutionary aspects of diet, theomega-6/omega-3ratio and genetic variation: nutritional implications for chronic diseases, *Biomedicine and Pharmacotherapy*, vol. 60, no. 9, pp. 502–507. Retrieved on May 18, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S0753332206002435>

- Smedes, F., & Askland, T. (1999). Revisiting the development of the Bligh and Dyer total lipid determination method. *Marine Pollution Bulletin*, 38(3), 193-201. Retrieved on May 18, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S0025326X98001702>
- Stansby, M. E. (1967). *Fish Oils: Their Chemistry, Technology, Stability, Nutritional Properties, and Uses*. M. E. Stansby (Ed.). Avi Publishing Company. Retrieved on May 18, 2017 from <http://www.sidalc.net/cgi-bin/wxis.exe/?IisScript=libri.xis&method=post&formato=2&cantidad=1&expresion=mfn=017787>
- Strlič, M., Cigić, I. K., Rabin, I., Kolar, J., Pihlar, B., & Cassar, M. (2009). Autoxidation of lipids in parchment. *Polymer degradation and stability*, 94(6), 886-890. Retrieved on May 18, 2017 from <https://www.sciencedirect.com/science/article/pii/S0141391009000901>
- Strocchi, A., & Bonaga, G. (1975). Correlation between urea inclusion compounds and conformational structure of unsaturated C18 fatty acid methyl esters. *Chemistry and Physics of Lipids*, 15(2), 87-94. Retrieved on May 18, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/000930847590033X>
- Suja, K. P., Abraham, J. T., Thamizh, S. N., Jayalekshmy, A., & Arumughan, C. (2004). Antioxidant efficacy of sesame cake extract in vegetable oil protection. *Food chemistry*, 84(3), 393-400. Retrieved on May 18, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S0308814603002486>
- Sun, Y., Kaksonen, M., Madden, D. T., Schekman, R., & Drubin, D. G. (2005). Interaction of Sla2p's ANTH domain with PtdIns (4, 5) P₂ is important for actin-dependent endocytic internalization. *Molecular biology of the cell*, 16(2), 717-730. Retrieved on June 18, 2017 from <https://www.molbiolcell.org/doi/full/10.1091/mbc.e04-08-0740>
- Tsai, C. E., Wooten, J. T., & Otto, D. A. (1989). Stability of fish oil in a purified diet with added antioxidants: Effects of temperature and light. *Nutrition Research*, 9(6), 673-678. Retrieved on May 18, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S0271531789801344>
- Uauy, R., & Valenzuela, A. (2000). Marine oils: the health benefits of n-3 fatty acids. *Nutrition (Burbank, Los Angeles County, Calif.)*, 16(7-8), 680-684.

Retrieved on May 18, 2017 from <https://pascal-francis.inist.fr/vibad/index.php?action=getRecordDetail&idt=1455867>

Undeland, I. (1999). Lipid oxidation in fillets of herring (*Clupea Harengus*) during processing and storage. *Lipidforum Nytt/News, Vår/Spring*, (57), 16-21. Retrieved on May 18, 2017 from <https://www.diva-portal.org/smash/record.jsf?pid=diva2%3A967048&dswid=-449>

Wang, C., Harris, W. S., Chung, M., Lichtenstein, A. H., Balk, E. M., Kupelnick, B., ... & Lau, J. (2006). n- 3 Fatty acids from fish or fish-oil supplements, but not α -linolenic acid, benefit cardiovascular disease outcomes in primary- and secondary-prevention studies: a systematic review. *The American journal of clinical nutrition*, 84(1), 5-17. Retrieved on April 8, 2017 from <https://academic.oup.com/ajcn/article/84/1/5/4633070?login=true>

Wanasundara, U. N., & Shahidi, F. (1998). Lipase-assisted concentration of n-3 polyunsaturated fatty acids in acylglycerols from marine oils. *Journal of the American Oil Chemists' Society*, 75(8), 945-951. Retrieved on April 8, 2017 from <https://aocs.onlinelibrary.wiley.com/doi/abs/10.1007/s11746-998-0271-9>

Wanasundara, U. N., & Shahidi, F. (1999). Concentration of omega 3-polyunsaturated fatty acids of seal blubber oil by urea complexation: optimization of reaction conditions. *Food Chemistry*, 65(1), 41-49. Retrieved on April 8, 2017 from <https://www.sciencedirect.com/science/article/abs/pii/S0308814698001538>

Zuta, C. P. (2003). Synthesis of novel triglycerides from mackerel by-products and vegetable oils (Doctoral dissertation, McGill University). Retrieved on April 1, 2017 from <https://www.collectionscanada.gc.ca/obj/thesescanada/vol2/QMM/TC-QMM-84863.pdf>

Gunning Fog Index: 9.44
Flesch Reading Ease: 54.24
Grammar Checking: 90/100
Plagiarism: 4%